



IO17 | Large Scale Bioinformatics for Immuno-Oncology

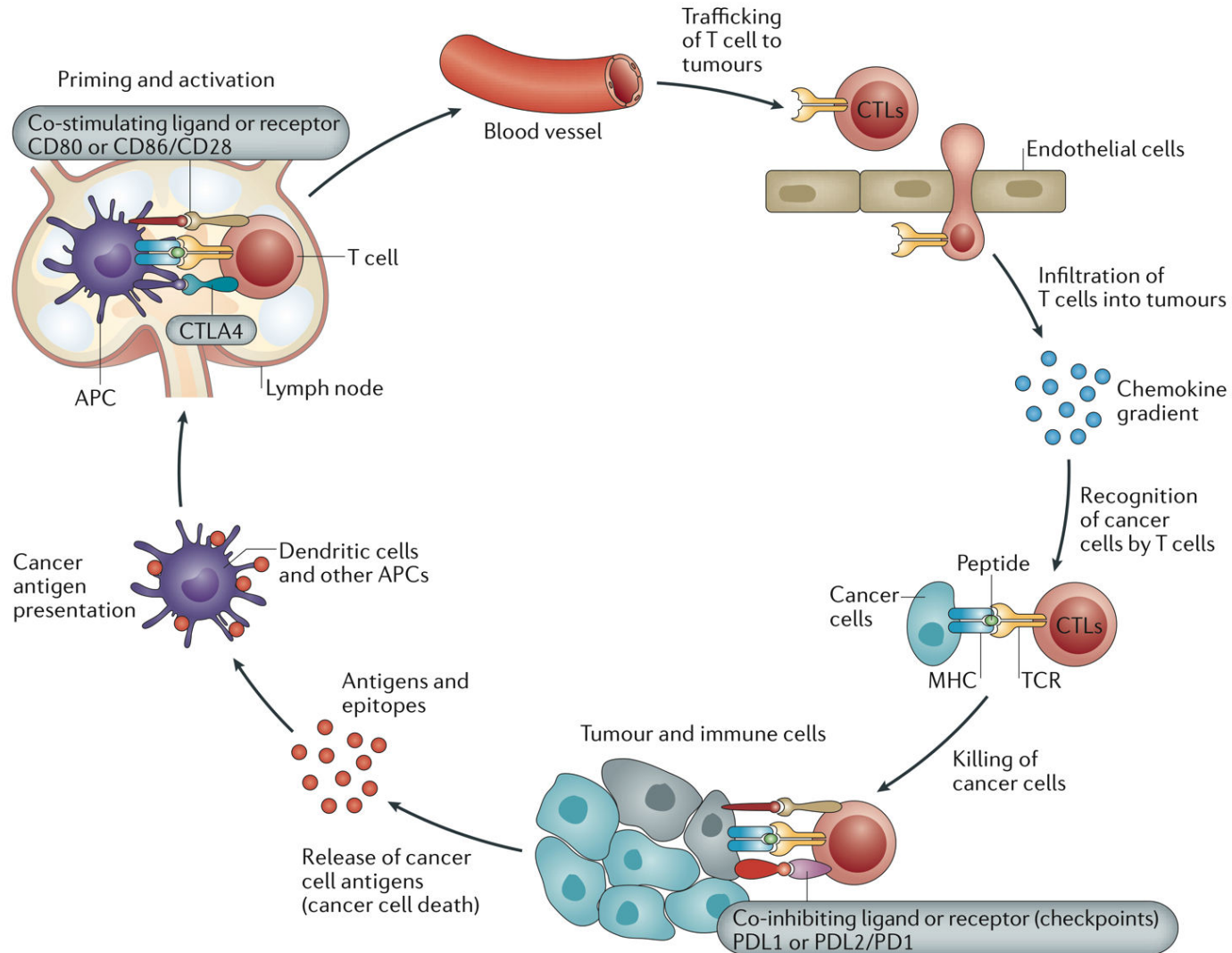
Prediction of tumor neoantigens

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The anticancer immune response

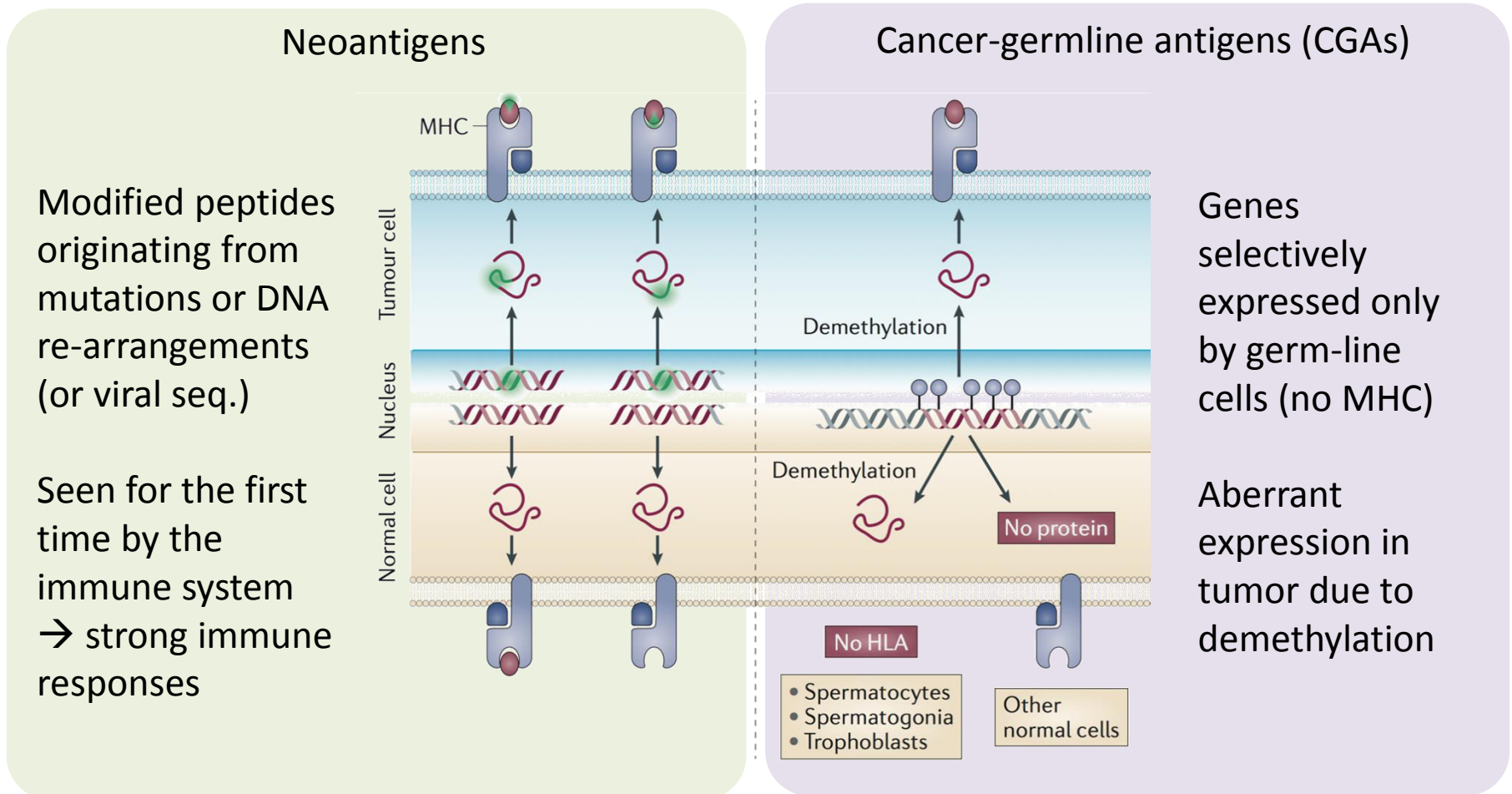
Effective anticancer immune responses require a series of stepwise events:



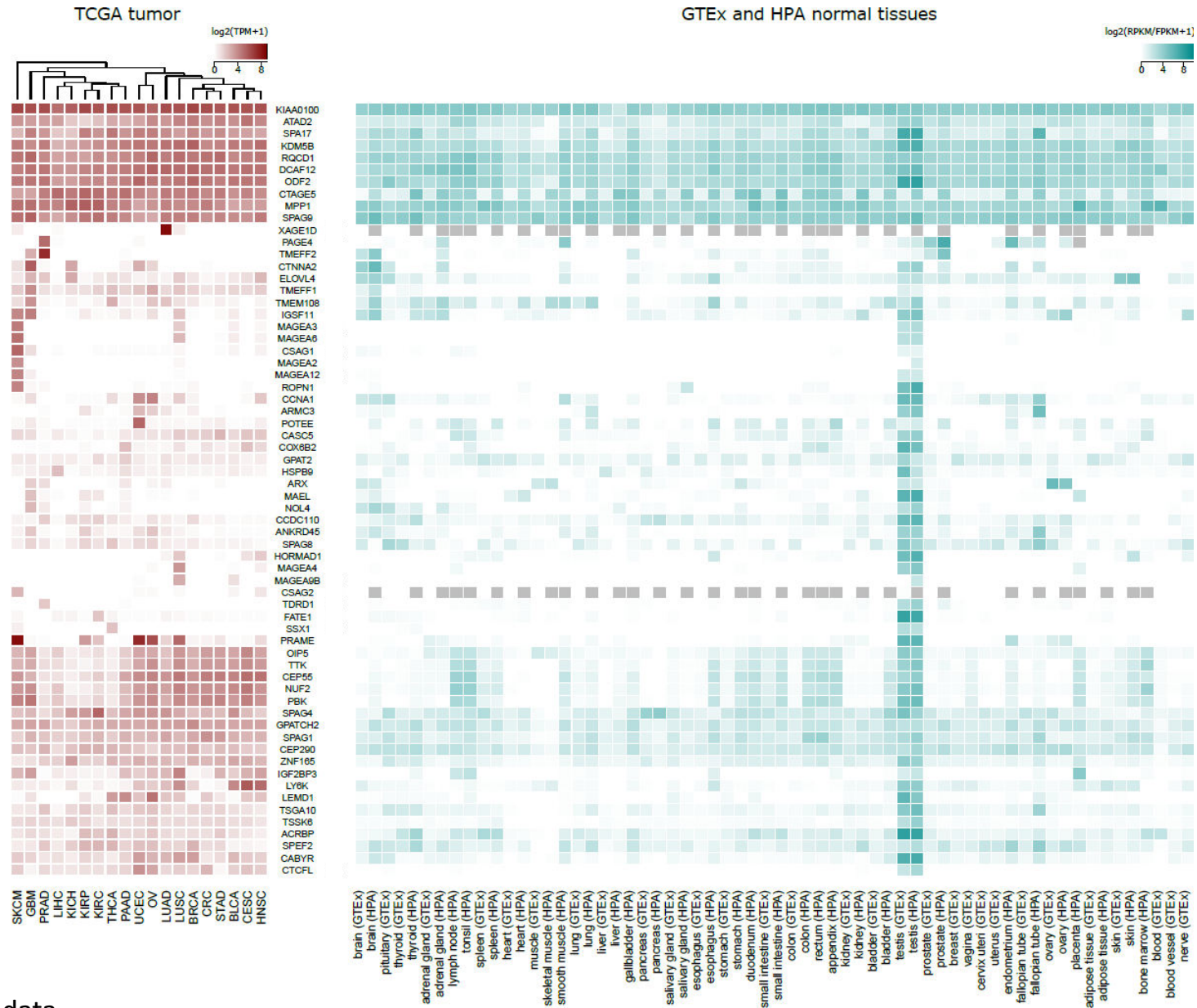
Tumor antigens

Tumor antigens are small peptides bound to the MHC molecules of tumor cells that can be recognized as “non-self” by the immune system

Only two classes of tumor antigens elicit immune responses that are strictly tumor specific:



Expression of cancer-germline antigens in solid tumors



Neoantigens in solid tumors

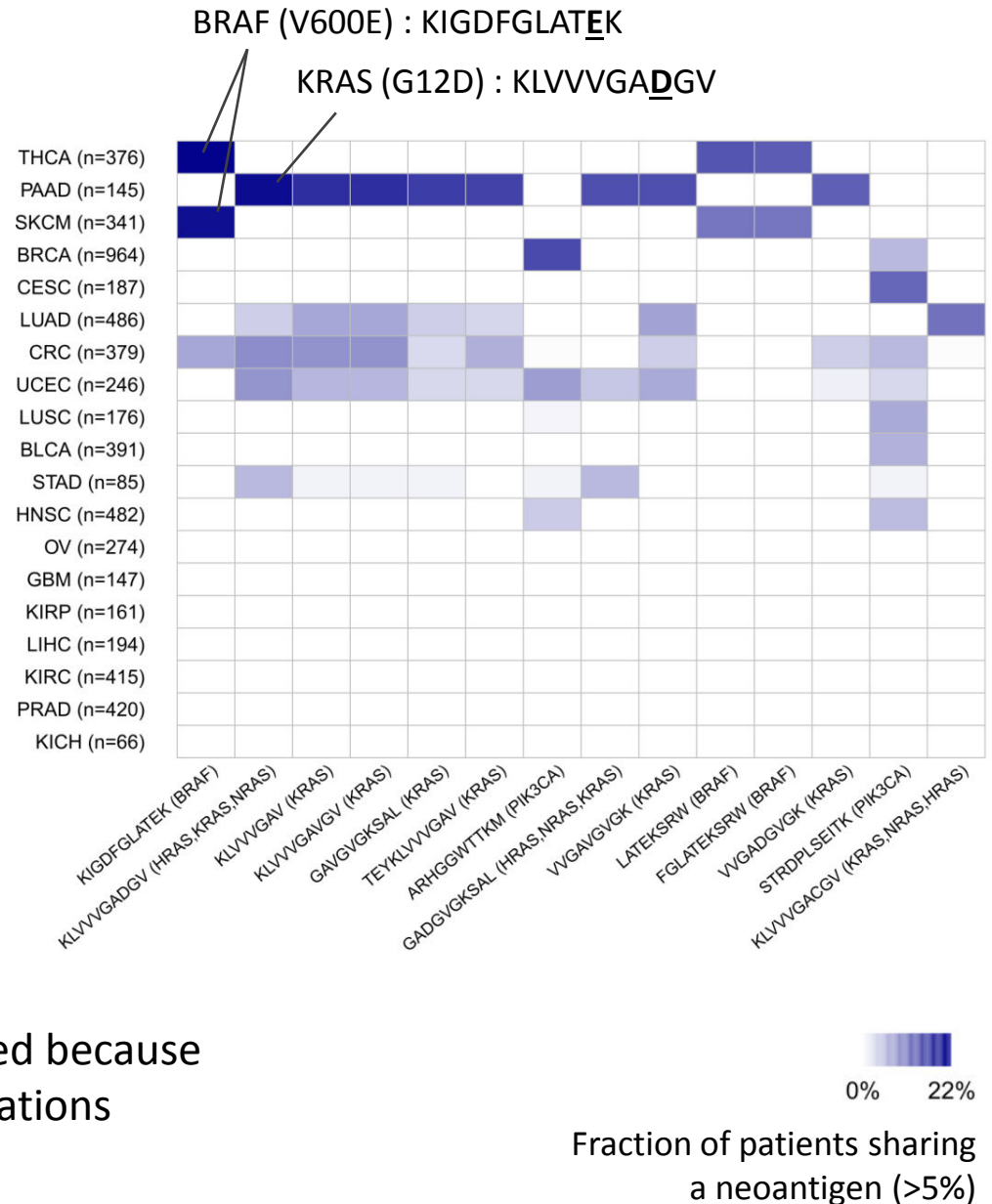
~1 petabyte of NGS data from
19 human cancers from TCGA
→ 933,954 neoantigens

7.0-10.6% of neoantigens
from driver mutations

Neoantigens shared in >15%
patients might be good
candidates for vaccination

Neoantigens are **diverse** and not shared because
they arise mainly from **passenger** mutations

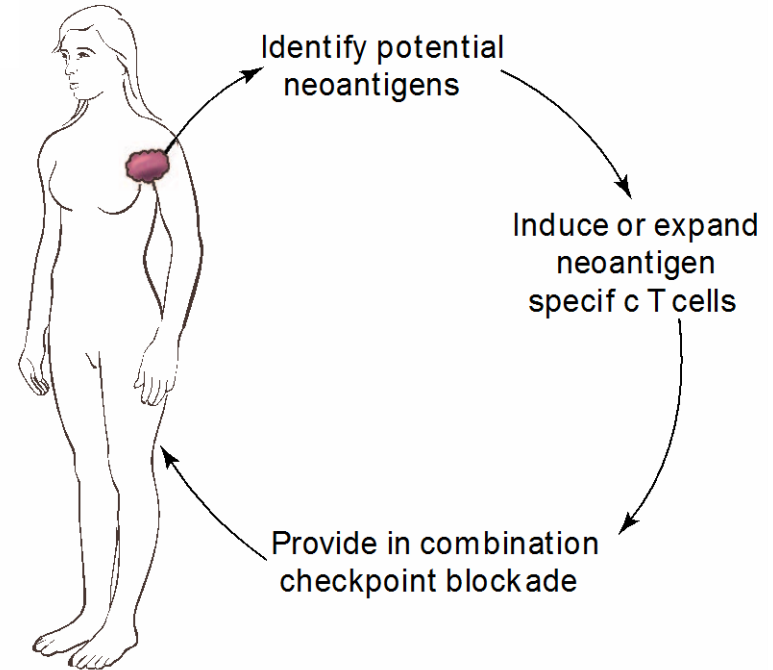
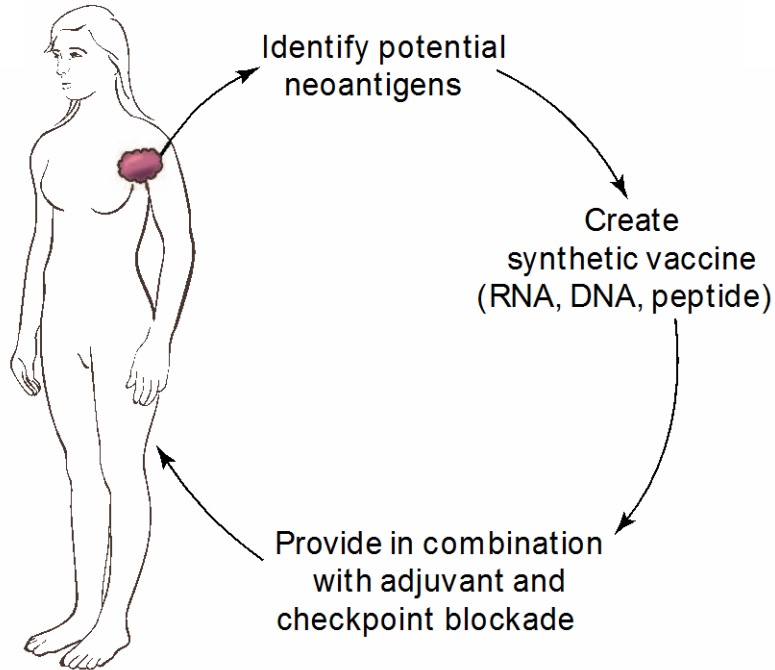
→ personalized therapy



Importance of tumor neoantigens for immunotherapy

- Immunotherapy has higher success rate in cancer types with high number of non-synonymous mutations (e.g. melanoma or MSI cancers)
- Mutations → neoantigens → recognized as non-self by T cells
- Mutations/neoantigens proposed as biomarker for immunotherapy
- But no clear-cut separation and some cancer types with low mutational load also respond (e.g. clear renal cell carcinoma)

Targeting patient-specific neoantigens



- Synthetic vaccine: DNA minicassette electroporated into patient-derived DCs, RNA vaccine, injected peptide vaccine
- Neoantigen-specific T cells: from patient or healthy donors

LETTER

doi: [10.1038/nature22991](https://doi.org/10.1038/nature22991)

An immunogenic personal neoantigen vaccine for patients with melanoma

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Synthetic long peptides representing up to 20 patient-specific neoantigens

LETTER

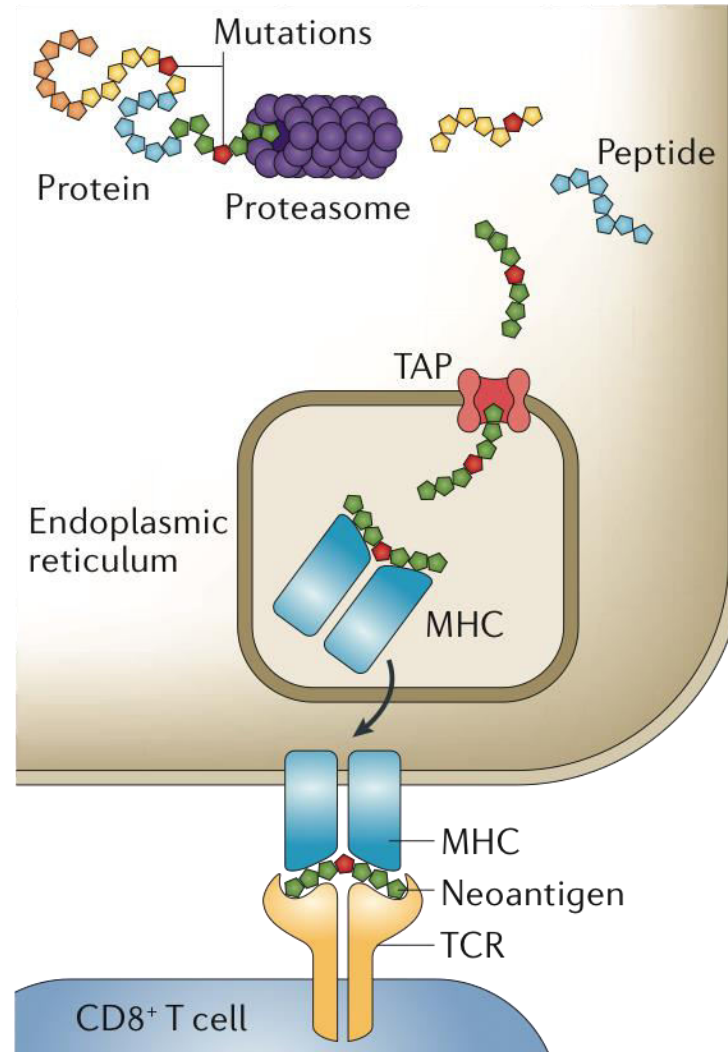
doi: [10.1038/nature23003](https://doi.org/10.1038/nature23003)

Personalized RNA mutanome vaccines mobilize poly-specific therapeutic immunity against cancer

Ugur Sahin^{1,2,3}, Evelyn Derhovanessian¹, Matthias Miller¹, Björn-Philipp Kloeke¹, Petra Simon¹, Martin Löwer², Valesca Bukur^{1,2}, Arbel D. Tadmor², Ulrich Luxemburger¹, Barbara Schrörs², Tana Omokoko¹, Mathias Vormehr^{1,3}, Christian Albrecht², Anna Paruzynski¹, Andreas N. Kuhn¹, Janina Buck¹, Sandra Heesch¹, Katharina H. Schreeb¹, Felicitas Müller¹, Inga Ortseifer¹, Isabel Vogler¹, Eva Godehardt¹, Sebastian Attig^{2,3}, Richard Rae², Andrea Breitzkreuz¹, Claudia Tolliver¹, Martin Suchan², Goran Martic², Alexander Hohberger³, Patrick Sorn², Jan Diekmann¹, Janko Ciesla⁴, Olga Waksman⁴, Alexandra-Kemmer Brück¹, Meike Witt¹, Martina Zillgen¹, Andree Rothermel², Barbara Kasemann², David Langer¹, Stefanie Bolte¹, Mustafa Diken^{1,2}, Sebastian Kreiter^{1,2}, Romina Nemecek⁵, Christoffer Gebhardt^{6,7}, Stephan Grabbe³, Christoph Höller⁵, Jochen Utikal^{6,7}, Christoph Huber^{1,2,3}, Carmen Loquai^{3*} & Özlem Türeci^{8*}

RNA-based vaccines

How neoantigens originate



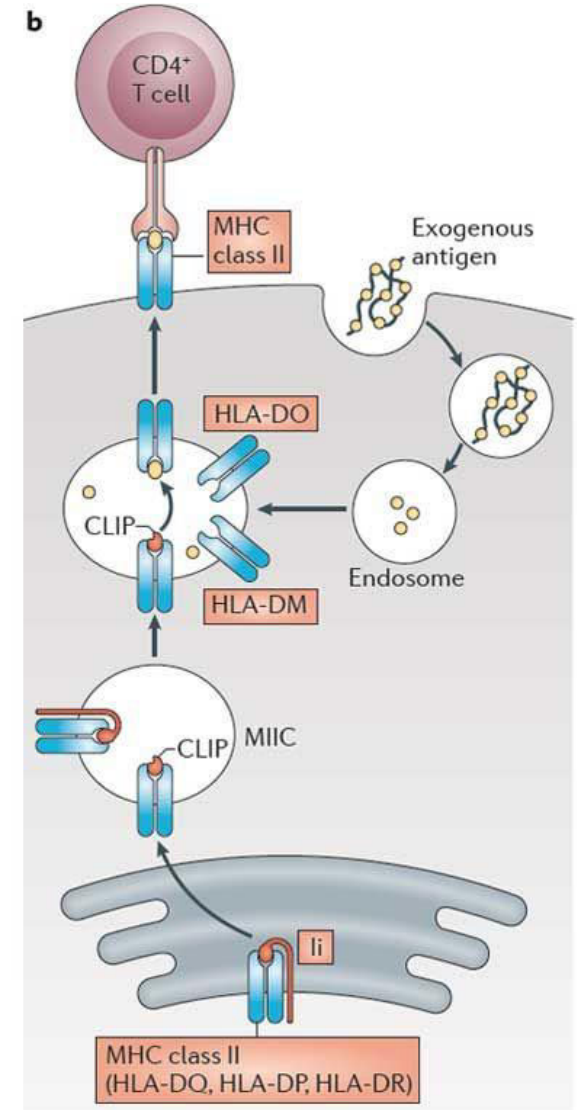
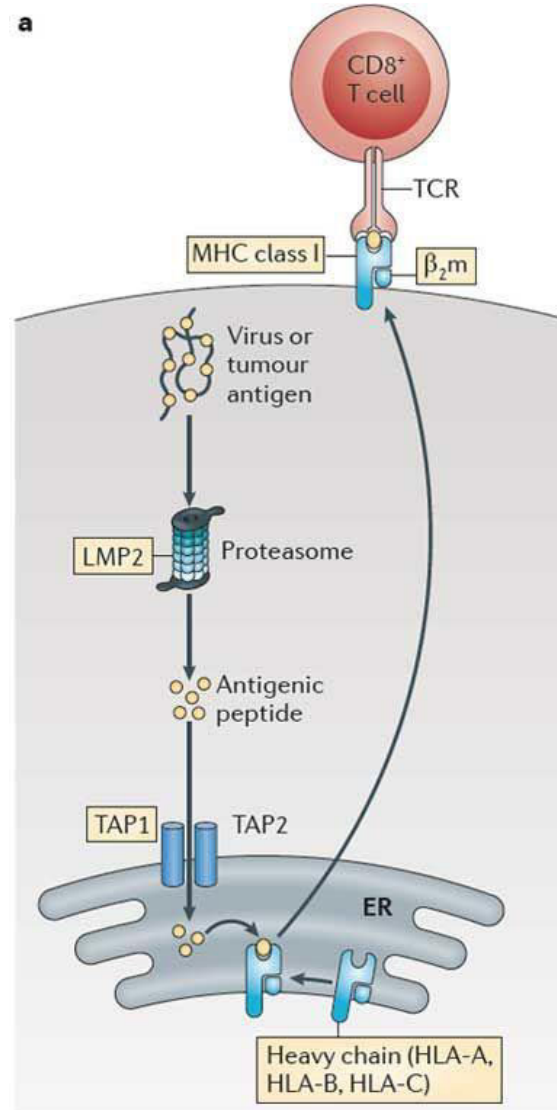
Class-I and class-II antigens

a) Class-I MHC molecules

- expressed on all nucleated cells (with some exceptions)
- Present 8-11 amino acid long peptides from intracellular proteins to **CD8+ T cells**

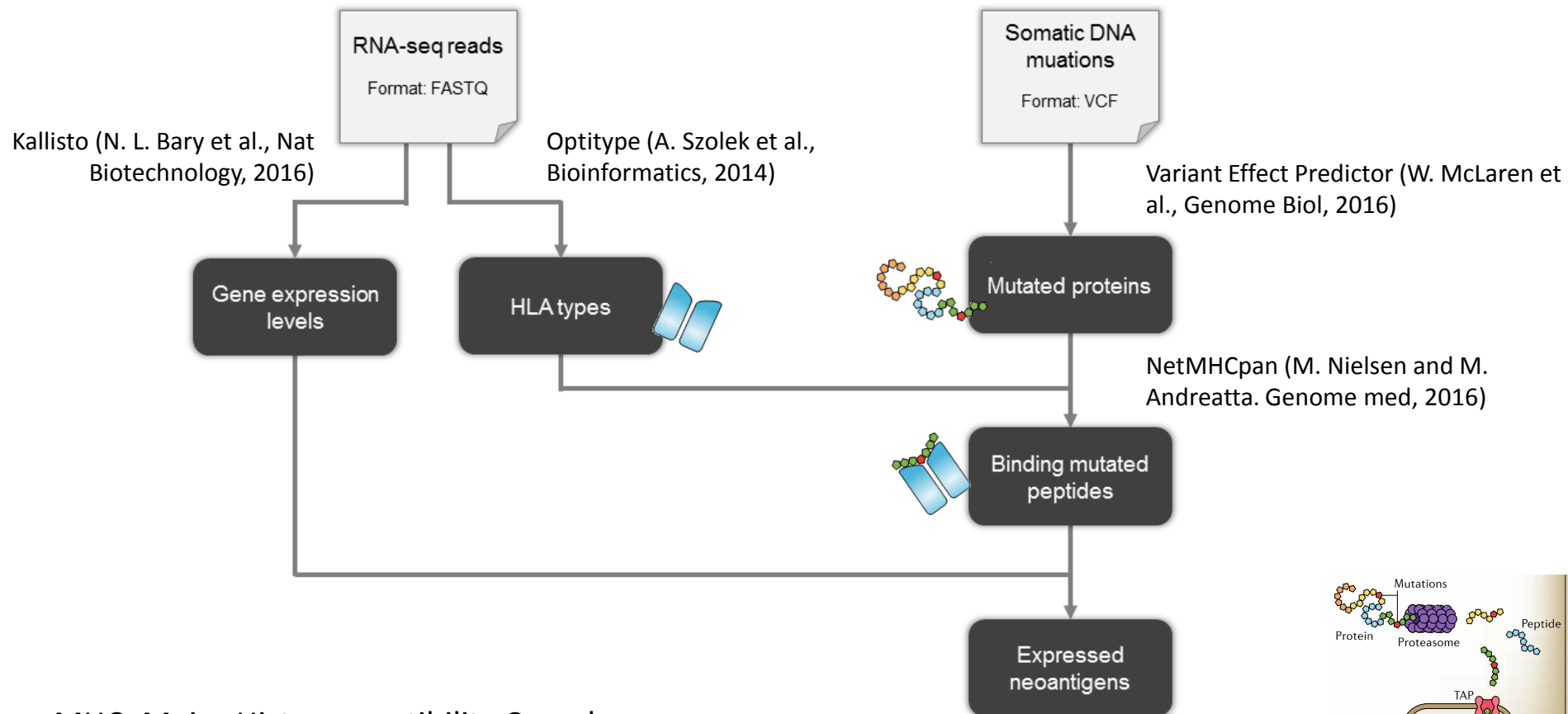
b) Class-II MHC molecules

- Expressed on professional antigen presenting cells (APC) like dendritic cells, macrophages, and B cells
- Present 10-30 amino acid long peptides from extracellular proteins to **CD4+ T cells**

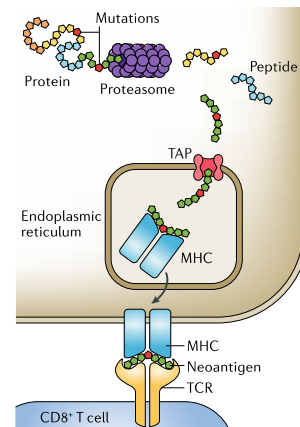


TIminer pipeline for the prediction of class-I neoantigen

Note: algorithms for class-I neoantigen prediction are more accurate than those for class-II



MHC: Major Histocompatibility Complex
HLA: Human Leukocyte Antigen (MHC in humans)



Running TIminer pipeline

TIminer is available as Docker image (easy installation and usage):

<http://www.icbi.at/software/timiner/timiner.shtml>

TIminer documentation:

<http://www.icbi.at/software/timiner/doc/index.html>

The full pipeline can be run with a single Python script: **TIminerPipeline.py**

After the installation, small example data can be analyzed with the full pipeline by executing from the “scripts” directory the command:

```
python TIminerPipeline.py --input ../samples/inputInfo.txt  
--out ../samples/out
```

Running single TIminer modules

```
from TIminer import TIminerAPI

TIminerAPI.executeKallisto(inputFile1="path/to/input_1.fastq",
                           inputFile2="path/to/input_2.fastq",
                           outputFile="path/to/norm_expr.txt",
                           subjectId="Subject_1",
                           threadCount=2)
```



Code saved in the script **myKallistoScript.py**

The Python script can be executed with:

```
python myKallistoScript.py
```



To use TIminer, Docker must be running on your computer!

Version for multi-sample analysis: `TIminerAPI.executeKallistoDir`